



qPCR SybrMaster UNG lowROX

Master mix for real-time qPCR with SYBR® Green fluorescent DNA stain

Cat. No.	Amount
PCR-376S	2 x 1,25 ml (2x conc.)
PCR-376L	10 x 1,25 ml (2x conc.)

For *in vitro* use only!

Shipping: shipped on gel packs

Storage Conditions: store at -20 °C

Additional Storage Conditions: avoid freeze/thaw cycles, store dark
Storage at 4 °C for up to 3 months possible.

Shelf Life: 12 months

Form: liquid

Concentration: 2x conc.

Spectroscopic Properties: λ_{exc} 494 nm (bound to DNA), λ_{em} 521 nm (bound to DNA)

Description:

qPCR SybrMaster UNG lowROX is designed for quantitative real-time analysis of DNA samples. The mix contains all reagents required for qPCR (except template and primers) in a premixed 2x concentrated ready-to-use solution. It is recommended for routine PCR applications, high throughput PCR or genotyping and provides an improved specificity and sensitivity when amplifying low-copy-number targets or working with complex backgrounds.

The mix is based on an optimized hot-start polymerase. Its activity is blocked by antibody at ambient temperature and switched on automatically at the onset of the initial denaturation. The thermal activation prevents the extension of nonspecifically annealed primers and primer-dimer formation at low temperatures during PCR setup.

The fluorescent DNA stain SYBR® Green intercalates into the amplification product during the PCR process and allows the direct quantification of target DNA without the need to synthesize sequence-specific labeled probes (i.g. TaqMan® Probes).

The mix contains UNG (Uracil-N-Glycosylase) and dUTP instead of dTTP to eliminate carry-over contamination of DNA from previous PCR reactions. The UNG treatment at the onset of thermal cycling removes uracil residues from dU-containing DNA and prevents it from serving as template.

The reaction chemistry of the kit is optimized for instruments that are compatible with the evaluation of a low ROX reference signal.

SYBR® Green Fluorescent DNA Stain

SYBR® Green Fluorescent DNA Stain is a superior DNA intercalator dye specially developed for DNA analysis applications including real-time PCR (qPCR). Upon binding to DNA, the non-fluorescent dye becomes highly fluorescent while showing no detectable inhibition to the PCR process. The dye is extremely stable, providing convenience during routine handling.

SYBR® Green is in contrast to EvaGreen® not recommended for high-resolution melting curve analysis (HRM).

To perform the SYBR® Green-based assay simply select the optical setting for SYBR® Green on the detection instrument.

ROX reference dye:

The qPCR SybrMaster with lowROX contains 50 nM ROX passive reference dye in the final assay. The dye does not take part in the PCR reaction but allows to normalize for non-PCR related signal variation and provides a baseline in multiplex reactions.

Content:

qPCR SybrMaster lowROX (red cap)

antibody-blocked hot start polymerase, UNG, dATP, dCTP, dGTP, dUTP, KCl, $(\text{NH}_4)_2\text{SO}_4$, MgCl_2 , SYBR® Green DNA intercalator dye, 100 nM ROX, additives and stabilizers

PCR-grade water

Preparation of the qPCR master mix:



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The preparation of a master mix is crucial in quantitative PCR reactions to reduce pipetting errors. Prepare a master mix of all components except template as specified. A reaction volume of 20-50 µl is recommended for most real-time instruments. Prepare 13 volumes of master mix for 12 samples or a triple-set of 4 samples. Pipet with sterile filter tips and minimize the exposure of the master mix to light. Perform the setup in an area separate from DNA preparation or analysis. No-template controls should be included in all amplifications.

component	20 µl assay	50 µl assay	final conc.
qPCR SybrMaster with UNG/lowROX	10 µl	25 µl	1x
primer forward (10 µM) ¹⁾	0.6 µl	1.5 µl	300 nM
primer reverse (10 µM) ¹⁾	0.6 µl	1.5 µl	300 nM
template DNA	x µl	x µl	-
PCR-grade water	fill up to 20 µl	fill up to 50 µl	-

¹⁾ The optimal concentration of each primer may vary from 100 to 500 nM.

Dispensing the master mix:

Vortex the master mix thoroughly to assure homogeneity and dispense the mix into real-time PCR tubes or wells of the PCR plate.

Addition of template DNA:

Add the remaining x µl of sample/template DNA to each reaction vessel containing the master mix and cap or seal the tubes/plate. Do not exceed 500 ng DNA per reaction as final concentration. Tubes or plates should be centrifuged before cycling to remove possible bubbles.

Recommended cycling conditions:

UNG treatment ²⁾	50 °C	2 min	1x
Initial denaturation and polymerase activation	95 °C	2 min	1x
Denaturation	95 °C	15 sec	35-45x
Annealing ³⁾	55-65 °C	20 sec	35-45x
Elongation ⁴⁾	72 °C	30 sec	35-45x

²⁾ Cycling step 1 is only required if an UNG (Uracil-N-Glycosylase) treatment is applied.

³⁾ The annealing temperature depends on the melting temperature of the primers used.

⁴⁾ The elongation time depends on the length of the amplicon. A time of 30 - 60 sec. for a fragment of up to 500 bp is recommended.

For optimal specificity and amplification an individual optimization of the recommended parameters, especially of the annealing temperature may be necessary for each new combination of template DNA and primer pair.

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Related Products:

Dual-labeled DNA probes